

SYNOPSIS

## DDT Strikes Back: Galapagos Sea Lions Face Increasing Health Risks

Juan José Alava, Sandie Salazar, Marilyn Cruz, Gustavo Jiménez-Uzcátegui,  
Stella Villegas-Amtmann, Diego Paéz-Rosas, Daniel P. Costa, Peter S. Ross,  
Michael G. Ikonomou, Frank A.P.C. Gobas

Received: 8 October 2010/Accepted: 8 January 2011/Published online: 18 February 2011

This synopsis was not peer reviewed.

### INTRODUCTION

Dichlorodiphenyltrichloroethane (DDT) is an effective and relatively cheap anti-malaria pesticide which has saved human lives, but also brought Northern wildlife species to the brink of extinction due to its persistence, bioaccumulation in the food-web, and hormone-mimicking characteristics (Carson 1962; Vallack et al. 1998; UNEP 2001; ATSDR 2002; Blus 2003; WHO 2006; Colborn and Carroll 2007; Van den Berg 2008). Thirty-two years after its ban in the US and its inclusion on Schedule 2 of the UN Stockholm Convention on Persistent Organic Pollutants (POPs) because of its damaging health effects in human and wildlife populations (UNEP 2001; ATSDR 2002; WHO 2006; Vallack et al. 1998; Colborn and Carroll 2007; Blus 2003), the WHO has recommitted to the use of the malaria-fighting pesticide DDT in tropical countries because of an increase in malaria cases in tropical countries (e. g., Peru, Bolivia, Paraguay, Brazil, and African nations), where DDT use was halted or low application rates were used (Van den Berg 2008; WHO 2006; Roberts et al. 1997, 2000).

Although evolutionary adaptation of the malaria vector has triggered insecticide resistance to DDT (Denholm et al. 2002), the return to an increased reliance on DDT use is testament to DDT's effectiveness in combating malaria at a low cost. Large volumes of DDT (i.e., 4,000–5,000 tonnes of active ingredient per year) were used in India and Africa countries to overcome the malaria crisis (Van den Berg 2008). However, 50 years of research on DDT in the

temperate and arctic regions of the Northern hemisphere have documented that human health benefits go hand in hand with detrimental effects on wildlife populations (Carson 1962; De Guise et al. 1998; Lahvis et al. 1995; Guillette et al. 1994; Blus 2003). Since wildlife provides life support, sustenance, and economic opportunities through harvesting and ecotourism in many developing tropical countries, the health of wildlife cannot be viewed in isolation of human health. A balance needs to be struck between health benefits and health risks of DDT use. This requires an understanding of the fate, trends, and dose-response relationship for DDT in tropical wildlife. Such information does currently not exist and monitoring and source control programs that can help to document this relationship are as absent in tropical countries to date as they were in the Western world when DDT was first used in the 1940s and 1960s (Roberts et al. 2000; ATSDR 2002; Blus 2003; Colborn and Carroll 2007; Van den Berg 2008).

From a global perspective, the protection of coastal food webs from contamination by chemical pollutants is critical to the long term conservation of the biodiversity and native inhabitants residing in unique places of the Earth, including UNESCO Global Heritage sites such as the Galapagos Islands. Coastal waters that are contaminated with persistent chemicals and pathogens can lead to human illness and adverse health, reduced fisheries quality and quantity, and impacts of the health of marine wildlife. This had obvious social and economic consequences. Conversely, coastal waters that are protected from chemical pollutants provide for an abundance of clean fisheries products and wildlife, and essential foundation for the well-being of the local biodiversity, human residents and the ecotourism sector.

At the top of the marine-coastal food chain, marine mammals can provide an “integrated” overview of ecosystem

health. As aquatic animals, they are also vulnerable to infection by pathogens of terrestrial origin. By documenting the presence of chemical pollutants in this species, we are able to deliver science-based advice to conservationists, managers, regulators, and stakeholders, on the implementation of best management practices. Equivalent to the role of killer whales as global sentinels of pollution in the Northeastern Pacific, the Galapagos sea lion has recently been used as a sentinel of marine pollution and a key indicator of the ecosystem functioning and oceanic-coastal environmental health in Galapagos Islands. Therefore, this synopsis aims to synthesize our recent findings on DDT contamination in Galapagos sea lions and its use as a sentinel-model species of environmental pollution by POPs.

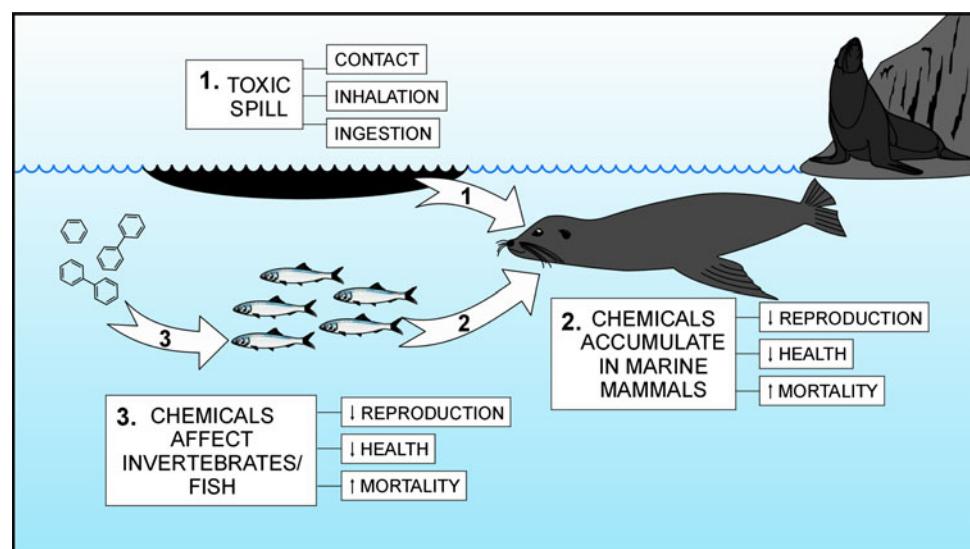
## DDT IN GALAPAGOS SEA LIONS

As most marine mammals, Galapagos sea lions do not escape from the global contamination by POPs and other chemical assaults, including oil spills (Fig. 1). Persistent organic pollutants have the capacity to bioaccumulate in marine food webs, resulting in the biomagnification of these contaminants in high trophic level predators such as marine mammals, as shown in Fig. 1. To start documenting the response of tropical wildlife to an increased use in DDT, we measured DDT concentrations in blubber biopsy samples collected from endemic and endangered Galapagos sea lion pups (*Zalophus wollebaeki*; Fig. 2) of the Galapagos Islands, located at 1,000 km offshore from continental Ecuador, in 2005 ( $n = 21$  pups) before the 2006 WHO recommendation and in 2008 ( $n = 20$  pups) after the WHO recommendation (Alava et al. 2011); we also conducted a risk assessment of the health impacts of DDT concentrations and its main metabolite *p,p'*-DDE by

comparing the detected concentrations in pups against toxic effect reference values documented in the literature as documented elsewhere (Alava 2011; Alava et al. 2011).

DDT mean  $\pm$  (SE) concentrations in samples collected in 2008 ( $530 \pm 110 \mu\text{g kg}^{-1}$  lipid) were significantly greater than concentrations ( $280 \pm 150 \mu\text{g kg}^{-1}$  lipid), measured in 2005 ( $t$  test = 3.465,  $P = 0.0013$ ), indicating a plausible increase of 86% in DDT levels (Alava et al. 2011). Based on the 2008 data (Alava et al. 2011), we found evidence that 1% of the *p,p'*-DDE concentrations exceeded the *p,p'*-DDE anti-androgenic effect reference values causing endocrine disruption in mammalian cell cultures (Kelce et al. 1995), 12% of the *p,p'*-DDE concentrations were above the minimum *p,p'*-DDE immunotoxic effect concentration reported for bottlenose dolphins (Lahvis et al. 1995). This indicates that DDT concentrations in Galapagos sea lion pups are near levels expected to be associated with impacts on the immune system. Synergistic and/or additive toxic effects due to mixtures with other pollutants with similar mode of toxicity can contribute to impacts on the immune and endocrine systems as concentrations of polychlorinated biphenyls (PCBs) and polybrominated diphenyl ether (PBDEs) flame retardants were also recently detected in these animals (Alava et al. 2009). POPs can bioaccumulate in oceanic and coastal-marine food chains of the Galapagos sea lion, compromising its health and reproductive status (Alava et al. 2010; Alava 2011; Alava et al. 2011). A compromised immune and endocrine system affects the ability of animals to combat disease and to successfully reproduce, which is especially relevant during periods of nutritional and other types of stress in stochastic environments (e.g. El Niño events) when mass mortality occurs and populations often approach the critical point of extinction (Alava and Salazar 2006; Vargas et al. 2007).

**Fig. 1** Galapagos sea lion can be exposed to oil spills, which can possess acute and chronic toxic effects, and Persistent Organic Pollutants (1), which can be accumulated mainly through dietary ingestion and by inhalation, causing potential health effects, including impairments to reproduction (2), due to contamination of diet items (fish preys) in the food chain (3). The prey can be also affected by contaminants (3). Adapted from Alava (2011)



**Fig. 2** The Galapagos sea lion is an endemic marine mammal to the Galapagos Islands and an endangered pinniped species, which have declined from 40,000 to 16,000 animals since the late 1970s (Picture: J.J. Alava)



## THE DDT LEGACY CONTINUES

The apparent increase of DDT levels in remote Galapagos sea lions is not an isolated event since concentrations of DDT in Adélie penguins (*Pygoscelis adeliae*) from remote areas of the western Antarctic Peninsula have not decreased between 2004 and 2006 (Geisz et al. 2008). Likewise, concentrations of DDT in human breast milk from Japan have not decreased since 1998 (Kunisue et al. 2006). DDT concentrations in Galapagos sea lion pups are lower than those detected in pinnipeds from the Northern Hemisphere (Kajiwara et al. 2001; Kannan et al. 2004; Debier et al. 2005; Del Toro et al. 2006; Blasius and Goodmanlowe 2008; Mos et al. 2010), but greater than those detected recently in adult subdominant males, adult females, juveniles and pups of southern elephant seals (*Mirounga leonina*) from Elephant Island, Antarctica (Miranda-Filho et al. 2007). Interestingly, Galapagos sea lion pups exhibited DDT concentrations similar to those detected in juveniles of Hawaiian monk seals (*Monachus schauinslandi*) from several subpopulations in the Northwestern Hawaiian Islands (Ylitalo et al. 2008). The maximum levels of DDT concentration (1,200–3,100 µg kg<sup>-1</sup> lipid) detected in six Galapagos sea lion pups are comparable to or higher than the concentrations recently found in Amazon River dolphins (*Inia geoffrensis*; 1,624 µg kg<sup>-1</sup> lipid) (Torres et al. 2009) and higher than concentrations measured in human milk (370 µg kg<sup>-1</sup> lipid) samples from the Brazilian Amazon (Azeredo et al. 2008), where DDT has been widely sprayed.

Although it is uncertain what the sources are from which DDT is delivered to the Galapagos, the long-range global atmospheric-oceanic transport (i.e. “multi-hopping effect”) and DDT revolatilization to the atmosphere (Iwata et al. 1994; Wilkering et al. 2000; Guglielmo et al. 2009) are likely the major mechanisms responsible for the incidence of DDT in the Galapagos. Recent modeling work reports that residence times and proportions of the total global masses of DDT are 10–15 days and 2% in the atmosphere, and 1.2 years and 26% in the global ocean with 30% of the DDT mass bounded to the organic matter phase in the equatorial Pacific Ocean, where high primary productivity exists due to nutrient-enriched waters (i.e., wind driven upwelling) (Guglielmo et al. 2009), as those found in Galapagos waters (Alava 2009). These observations portray that the physical-chemical properties of DDT, oceanographic conditions, and chronic atmospheric inputs are the driving forces explaining the presence of DDT in wild life of the islands.

## CONSERVATION AND MANAGEMENT IMPLICATIONS

The Galapagos is one of the last living natural labs to study evolution and was already a UNESCO-Heritage site at risk due to invasive species, escalating human population growth and burgeoning tourism (Watkins and Cruz 2007), but the impact by persistent organic pollutants, including DDT, has been scarcely studied (Alava and Salazar 2006;

Alava et al. 2009). While additional temporal monitoring of DDT levels in Galapagos sea lions will provide greater insight into environmental responses to recent DDT use, our study provides a timely warning signal of the dangers of an increased reliance of DDT for malaria control in tropical countries. DDT is not presently used in the Galapagos Islands, underscoring the ready transport away from applications zones, and emphasizing the vulnerability of remote tropical regions. The rise of DDT concentrations in tropical wild life species is of great concern because of the well known toxicity of DDT and the demonstrated effects of DDT on bird (Hickey and Anderson 1968; Blus 2003) and other wildlife species in the Northern hemisphere (Guillette et al. 1994; De Guise et al. 1998; Vallack et al. 1998; Ylitalo et al. 2005). To ensure that tropical wildlife species will not meet a similar fate in the future, it is important that the DDT issue is brought to global attention immediately as well as the urgent need of environmental monitoring programs.

Here, we show that DDT concentrations in endangered Galapagos sea lion pups increased significantly between 2005 and 2008 and have attained levels that can affect the immune system and population dynamics during periods of nutritional stress (El Niño events). Concentrations of DDT and associated health risks in wildlife are generally believed to be declining but this may no longer be the case in tropical countries where DDT is increasingly used. The toxicological principle “*the dose makes the poison*” provides the scientific rationale for benefiting from the anti-malaria properties of DDT while minimizing or avoiding damage to local and global wildlife and advocates the implementation of source control. However, its precautionary application requires a commitment to monitoring remote environments, including the much-valued Galapagos Island Archipelago, and impact assessment and source control programs that have received scant attention in developing nations to date.

**Acknowledgments** We are grateful to the volunteers from the Marine Mammal Center in Santa Barbara (CA, USA) for supporting the cruise expedition and their expertise in sea lion capture. We thank P. Martinez, A. Paras, G. Merlen, and the Galapagos National Park rangers for their assistance during the sampling. The field sampling was possible thanks to the Project Health Status, Genetic and Rescue Techniques of Galapagos Pinnipeds of the Charles Darwin Foundation, and the Galapagos National Park Service. This work is part of the contribution number 2007 of the Charles Darwin Foundation. The export permit of the samples was authorized by the Galapagos National Park.

## REFERENCES

- Alava, J.J., and S. Salazar. 2006. Status and conservation of Otariids in Ecuador and the Galapagos Islands. In *Sea lions of the world*, ed. A.W. Trites, S.K. Atkinson, D.P. DeMaster, L.W. Fritz, T.S. Gelatt, L. D. Rea, and K.M. Wynne, 495–519. Fairbanks: Alaska Sea Grant College Program, University of Alaska Fairbanks.
- Alava, J.J. 2009. Carbon productivity and flux in the marine ecosystems of the Galapagos Marine Reserve based on cetacean abundances and trophic indices. *Revista de Biología Marina y Oceanografía* 44: 109–122.
- Alava, J.J., S. Salazar, G. Jiménez-Uzcategui, M. Cruz, D. Páez-Rosas, P. Calle, J. Denkinger, P.S. Ross, M. Ikonomou, and F. Gobas. 2010. Biomagnificación de Contaminantes Orgánicos Persistentes (COPs) en el Lobo Marino de Galápagos (*Zalophus wollebaeki*): Implicaciones para la conservación de la Biodiversidad Marina. 1er Simposio Latinoamericano de Biodiversidad Marina y Costera de Latinoamérica y El Caribe, y 2do Simposio Ecuatoriano de Biodiversidad Marino Costera. Ministerio del Ambiente, Ecuador; Grupo de Trabajo sobre Biodiversidad Marino y Costera del Ecuador. 9–12 Diciembre, 2010, Manta, Ecuador.
- Alava, J.J. 2011. Bioaccumulation of pollutants in Galapagos sea lions and marine mammals from British Columbia, Canada. PhD Thesis, School of Resource and Environmental Management, Faculty of Environment, Simon Fraser University, BC, Canada.
- Alava, J.J., M.G. Ikonomou, P.S. Ross, D. Costa, S. Salazar, and F.A.P.C. Gobas. 2009. Polychlorinated biphenyls (PCBs) and polybrominated diphenyl ethers (PBDEs) in Galapagos sea lions (*Zalophus wollebaeki*). *Environmental Toxicology and Chemistry* 28: 2271–2282.
- Alava, J.J., P.S. Ross, M.G., Ikonomou, M. Cruz, G. Jimenez-Uzcategui, S. Salazar, D.P. Costa, S. Villegas-Amtmann, P. Howorth, and F.A.P.C. Gobas. 2011. DDT in endangered Galapagos Sea Lions (*Zalophus wollebaeki*) from eight rookeries in the Galapagos Islands. *Marine Pollution Bulletin* (accepted).
- ATSDR (Agency for Toxic Substances and Disease Registry). 2002. *Toxicological profile for DDT, DDE, DDD*, 403. Atlanta, Georgia: Division of Toxicology/Toxicology Information Branch, U.S. Department of Health and Human Services, Public Health Service.
- Azeredo, A., J.P.M. Torres, M. de Freitas Fonseca, J.L. Britto, W.R. Bastos, C.E. Azevedo e Silva, G. Cavalcanti, R.O. Meire, et al. 2008. DDT and its metabolites in breast milk from the Madeira River basin in the Amazon, Brazil. *Chemosphere* 73: S246–S251.
- Blasius, M.E., and G.D. Goodmanlowe. 2008. Contaminants still high into-level carnivores in the Southern California Bight: Levels of DDT and PCBs in resident and transient pinnipeds. *Marine Pollution Bulletin* 56: 1973–1982.
- Blus, L.J. 2003. Organochlorine pesticides. In *Handbook of ecotoxicology*, ed. D.J. Hoffman, B.A. Rattner, G.A. Burton, and J. Cairns, 313–339. Boca Raton: CRC Press.
- Carson, R. 1962. *Silent spring*. Boston: Houghton Mifflin Company.
- Colborn, T., and L.E. Carroll. 2007. Pesticides, sexual development, reproduction, and fertility: Current perspective and future direction. *Human and Ecological Risk Assessment* 13: 1078–1110.
- De Guise, S., D. Martineau, P. Beland, and M. Fournier. 1998. Effects of in vitro exposure of beluga whale leukocytes to selected organochlorines. *Journal of Toxicology and Environmental Health, Part A* 55: 479–493.
- Debier, C., G.M. Ylitalo, M. Weisse, F. Gulland, D.P. Costa, B.J. Le Boeuf, T. de Tillesse, and Y. Larondelle. 2005. PCBs and DDT in the serum of juvenile California sea lions: Associations with vitamins A and E and thyroid hormones. *Environmental Pollution* 134: 323–332.
- Del Toro, L., G. Heckel, V.F. Camacho-Ibar, and Y. Schramm. 2006. California sea lions (*Zalophus californianus californianus*) have lower chlorinated hydrocarbon contents in northern Baja California, Mexico, than in California, USA. *Environmental Pollution* 142: 83–92.

- Denholm, I., G.J. Devine, and M.S. Williamson. 2002. Insecticide resistance on the move. *Science* 297: 2222–2223.
- Geisz, H.N., R.M. Dickhut, M.A. Cochran, W.R. Fraser, and H.W. Ducklow. 2008. Melting glaciers: A probable source of DDT to the Antarctic Marine Ecosystem. *Environmental Science and Technology* 42: 3958–3962.
- Guglielmo, F., G. Lammel, and E. Maier-Reimer. 2009. Global environmental cycling of gamma-HCH and DDT in the 1980s—a study using a coupled atmosphere and ocean general circulation model. *Chemosphere* 76: 1509–1517.
- Guillette Jr., L. J., T.S. Gross, G.R. Masson, J.M. Matter, H.F. Percival, and A.R. Woodward. 1994. Developmental abnormalities of the gonad and abnormal sex hormone concentrations in juvenile alligators from contaminated and control lakes in Florida. *Environmental Health Perspectives* 102: 680–688.
- Hickey, J.J., and D.W. Anderson. 1968. Chlorinated hydrocarbons and eggshell changes in raptorial and fish-eating birds. *Science* 162: 271–273.
- Iwata, H., S. Tanabe, N. Sakai, A. Nishimura, and R. Tatsukawa. 1994. Geographical distribution of persistent organochlorines in air, water and sediments from Asia and Oceania, and their implications for global redistribution from lower latitudes. *Environmental Pollution* 85: 15–33.
- Kajiwara, N., K. Kannan, M. Muraoka, M. Watanabe, S. Takahashi, F. Gulland, H. Olsen, A.L. Blankenship, P.D. Jones, S. Tanabe, and J.P. Giesy. 2001. Organochlorine pesticides, polychlorinated biphenyls, and butyltin compounds in blubber and livers of stranded California sea lions, Elephant Seals, and harbor seals from coastal California, USA. *Archives of Environmental Contamination and Toxicology* 41: 90–99.
- Kannan, K., N. Kajiwara, B.J. Le Boeuf, and S. Tanabe. 2004. Organochlorine pesticides and polychlorinated biphenyls in California sea lions. *Environmental Pollution* 131: 425–434.
- Kelce, W.R., C.R. Stone, S.C. Laws, L.E. Gray, J.A. Kemppainen, and E.M. Wilson. 1995. Persistent DDT metabolite *p,p*-DDE is a potent androgen receptor antagonist. *Nature* 375: 581–585.
- Kunisue, T., M. Muraoka, M. Ohtake, A. Sudaryanto, N.H. Minh, D. Ueno, Y. Higaki, M. Ochi, et al. 2006. Contamination status of persistent organochlorines in human breast milk from Japan: Recent levels and temporal trend. *Chemosphere* 64: 1601–1608.
- Lahvis, G.P., R.S. Wells, D.W. Kuehl, J.L. Stewart, H.L. Rhinehart, and C.S. Via. 1995. Decreased lymphocyte responses in free-ranging bottlenose dolphins (*Tursiops truncatus*) are associated with increased concentrations of PCBs and DDT in peripheral blood. *Environ. Health Perspectives* 103: 67–72.
- Miranda-Filho, K.C., T.L. Metcalfe, C.D. Metcalfe, R.B. Metcalfe, R.B. Robaldo, M.M.C. Muelbert, E.P. Colares, P.E. Martinez, and A. Bianchini. 2007. Residues of persistent organochlorine contaminants in Southern Elephant Seals (*Mirounga leonina*) from Elephant Island, Antarctica. *Environmental Science and Technology* 41: 3829–3835.
- Mos, L., M. Cameron, S.J. Jeffries, B.F. Koop, and P.S. Ross. 2010. Risk-based analysis of PCB toxicity in harbor seals. *Integrated Environmental Assessment and Management* 6: 631–640.
- Roberts, D.R., L.L. Laughlin, P. Hsueh, and L.I.J. Legters. 1997. DDT, global strategies, and a malaria control crisis in South America. *Emerging Infectious Diseases* 3: 295–302.
- Roberts, D.R., S. Manguin, and J. Mouchet. 2000. DDT house spraying and re-emerging malaria. *Lancet* 356: 330–332.
- Torres, J.P.M., J. Lailson-Brito, G.C. Saldanha, P. Dorneles, C.E.A. Silva, O. Malm, J.R. Guimarães, A. Azeredo, et al. 2009. Persistent toxic substances in the Brazilian Amazon: Contamination of man and the environment. *Journal of the Brazilian Chemical Society* 20: 1175–1179.
- UNEP (United Nations Environment Program). 2001. *Final act of the conference of plenipotentiaries on the Stockholm convention on persistent organic pollutants*, 44. Stockholm, Sweden/Geneva, Switzerland: UNEP/POPS/CONF/4UNEP.
- Vallack, H.W., D.J. Bakker, I. Brandt, E. Broström-Lundén, A. Brouwer, K.R. Bull, C. Gough, R. Guardans, et al. 1998. Controlling persistent organic pollutants—what next? *Environmental Toxicology and Pharmacology* 6: 143–175.
- Van den Berg, H. 2008. *Global status of DDT and its alternatives for use in vector control to prevent disease*, 31. Geneva, Switzerland: Stockholm Convention on Persistent Organic Pollutants, United Nations Environment Programme, UNEP/POPS/DDTBP.1/2.
- Vargas, F.H., R.C. Lacy, P.J. Johnson, A. Steinfurth, R.J.M. Crawford, P.D. Boersma, and D.W. Macdonald. 2007. Modelling the effect of El Niño on the persistence of small populations: The Galápagos penguin as a case study. *Biological Conservation* 137: 138–148.
- Watkins, G., and F. Cruz. 2007. *Galapagos at risk: A socioeconomic analysis of the situation in the Archipelago*, 21. Puerto Ayora, Province of Galapagos, Ecuador: Charles Darwin Foundation.
- WHO (World Health Organization). 2006. *WHO gives indoor use of DDT a clean bill of health for controlling malaria: WHO promotes indoor residual spraying with insecticides as one of three main interventions to fight malaria*. 15 September 2006. World Health Organization, Washington, DC. <http://www.who.int/mediacentre/news/releases/2006/pr50/en/index.html>. Accessed 19 Oct 2008.
- Wilkening, K.E., L.A. Barrie, and M. Engle. 2000. Trans-Pacific air pollution. *Science* 290: 65–66.
- Ylitalo, G.M., J.E. Stein, T. Hom, L.L. Johnson, K.L. Tilbury, A.J. Hall, T. Rowles, D. Greig, L.J. Lowenstein, and F.M.D. Gulland. 2005. The role of organochlorines in cancer-associated mortality in California sea lions (*Zalophus californianus*). *Marine Pollution Bulletin* 50: 30–39.
- Ylitalo, G.M., M. Myers, B.S. Stewart, P.K. Yochem, R. Braun, L. Kashinsky, D. Boyd, G.A. Antonelis, S. Atkinson, A.A. Aguirre, and M.M. Krahn. 2008. Organochlorine contaminants in endangered Hawaiian monk seals from four subpopulations in the Northwestern Hawaiian Islands. *Marine Pollution Bulletin* 56: 231–244.

#### Juan José Alava (✉)

*Address:* School of Resource & Environmental Management, Simon Fraser University 8888, University Drive, Burnaby, BC V5A 1S6, Canada.

*Address:* Fundacion Ecuatoriana para el Estudio de Mamíferos Marinos (FEMM), Guayaquil, Ecuador.  
e-mail: jalavasa@sfu.ca

#### Sandie Salazar

*Address:* Charles Darwin Foundation, Puerto Ayora, Santa Cruz, Galápagos, P.O. Box 17-1-3891, Quito, Ecuador.

#### Marilyn Cruz

*Address:* Galápagos Genetics Epidemiology and Pathology Laboratory (GGEPL), Puerto Ayora, Galápagos Islands, Ecuador.

#### Gustavo Jiménez-Uzcátegui

*Address:* Charles Darwin Foundation, Puerto Ayora, Santa Cruz, Galápagos, P.O. Box 17-1-3891, Quito, Ecuador.

#### Stella Villegas-Amtmann

*Address:* Department of Ecology and Evolutionary Biology, University of California, 100 Shaffer Road, Santa Cruz 95060, CA, USA.

#### Diego Paéz-Rosas

*Address:* Centro Interdisciplinario de Ciencias Marinas, Instituto Politécnico Nacional, Ave. IPN s/n. Colonia Playa Palo de Santa Rita, La Paz, Baja California Sur C. P. 23060, Mexico.

**Daniel P. Costa**

*Address:* Department of Ecology and Evolutionary Biology, University of California, 100 Shaffer Road, Santa Cruz 95060, CA, USA.

**Peter S. Ross**

*Address:* Institute of Ocean Sciences, Fisheries and Oceans Canada, 9860 West Saanich Rd, Sidney, BC V8L 4B2, Canada.

**Michael G. Ikonomou**

*Address:* Institute of Ocean Sciences, Fisheries and Oceans Canada, 9860 West Saanich Rd, Sidney, BC V8L 4B2, Canada.

**Frank A.P.C. Gobas**

*Address:* School of Resource & Environmental Management, Simon Fraser University 8888, University Drive, Burnaby, BC V5A 1S6, Canada.